mRNA Purification and Analysis

mRNA HPLC Analysis on SEC, RP, Affinity, and IEX

Sepax Technologies, Inc.

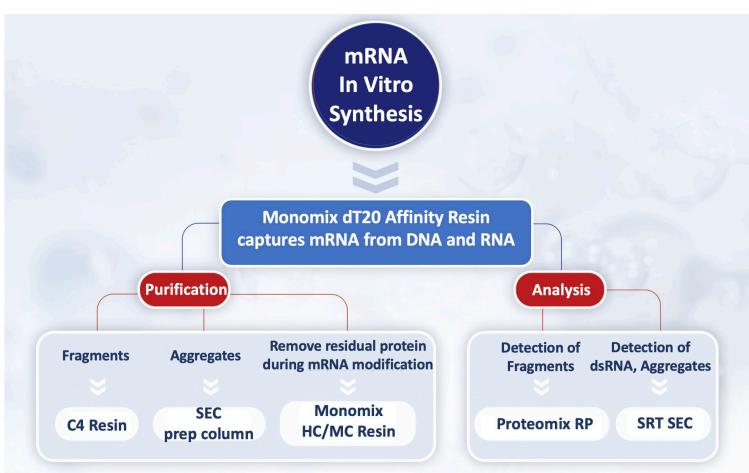
sepax

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Better Surface Chemistry for Better Separation

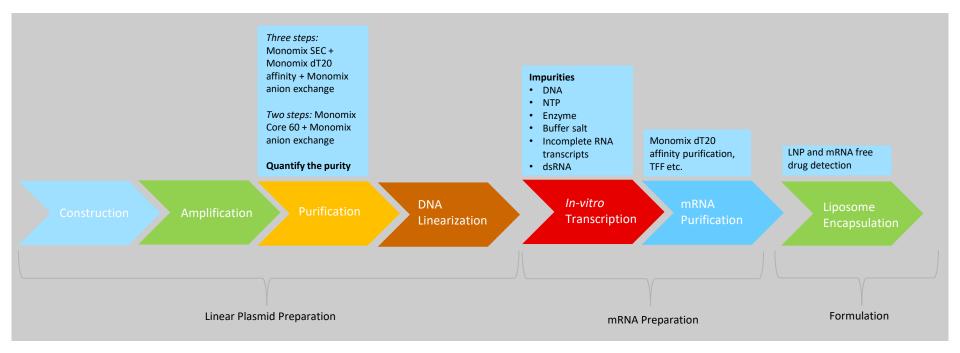
mRNA Chromatography Applications

The application of mRNA in the therapeutic and vaccine fields has led to a worldwide demand in chromatography technology for both analytical and manufacturing processes. mRNA drug and research development require robust and competent methods to evaluate mRNA integrity, which is essential for therapeutic effect and/or immunogenicity.





mRNA Production Process Example: Covid-19 mRNA Vaccine



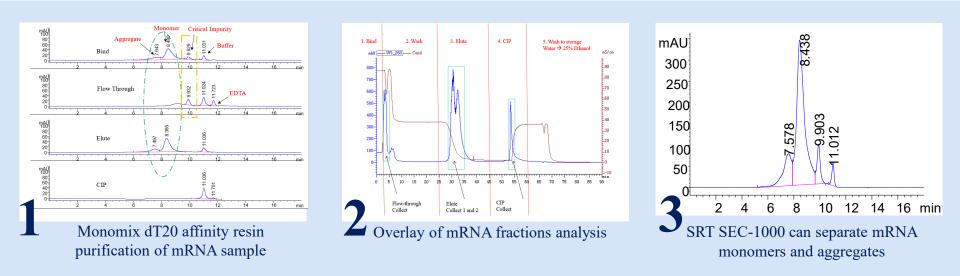


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General Description

Sepax Monomix dT20 Affinity Resin is 30 µm narrowly dispersed polymethacrylate rigid base bead is functionalized with a polyhydroxylated surface coating layer that provides a bioinert surface and shows low non-specific binding. Resin surface of Monomix dT20 can capture mRNA through base pairing between oligo dT20-mer and the mRNA polyA tail, which can be utilized to simplify the subsequent purification steps and maximize overall production efficiency. After purification, quality consistency of purified mRNA was accessed by Size exclusion chromatography SEC method, here using Sepax SRT SEC-1000 column. Slide 1 shows the complete running sequence (including Bind, Wash, Elute, CIP and Storage stages) of a simplified purification process of mRNA using Monomix dT20 affinity resin. Slide 2 shows the stack SEC profiles of initial mRNA sample (a) and the fractions collected from Wash (b), Elute (c), and CIP (d) stages on SRT SEC-1000 column.

In mRNA therapeutic development, size exclusion chromatography SEC has been utilized for its quick reliable method in many applications such as purification or aggregate quantification. Slide 3 shows the application of separating single stranded mRNA monomer, its aggregates including possible double stranded mRNA and mononucleotides. Additionally, Reverse-phase (RP) chromatography can also be used as an orthogonal method for further characterization of each SEC peak.





Sepax Monomix dT20 Affinity Resin



Part Number: 283030950-0000: 1 mL, 5 mL resin Part Number: 283030950-750100: 4.2 mL cartridge Part Number: 283030950P-2105: 2.1 x 50 mm PEEK Part Number: 283030950P-4605: 4.6 x 50 mm PEEK

Shipping and Storage Solvent: 20% Ethanol with Water Shipping Condition: Wet Ice



Characteristics

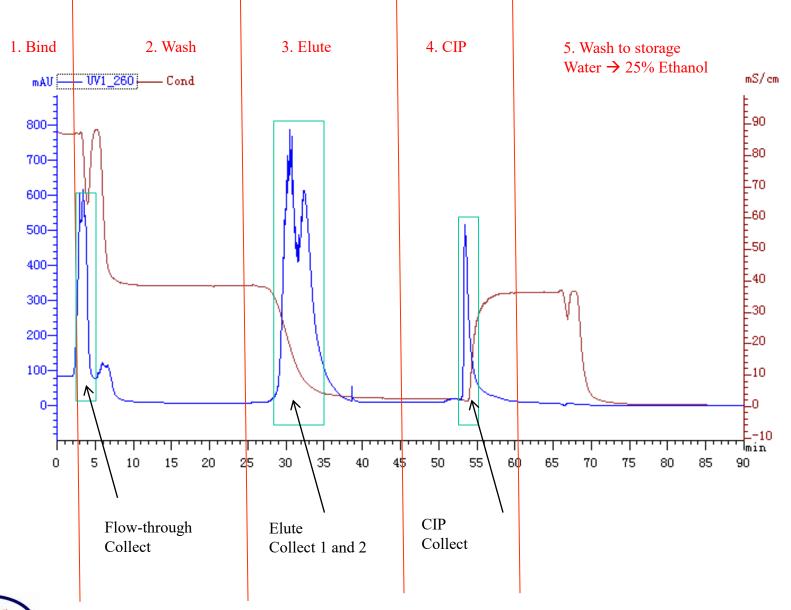
- Provides efficient capture and release under standard mRNA purification conditions, simplify subsequent purification steps and maximize overall production efficiency.
- Decreases process development time and enhances productivity.
- Remove plasmid DNA and other transcription components.
- Stable at elevated temperatures for the breakdown of undesired higher order structures, if needed.
- Excellent scalability. Provide prepacked columns, semi-prep columns, prep columns and bulk resin.
- Non-animal derived
- Resin customization. Polymethacrylate bead size is available at 10, 15, 30 and 60μm.



Technical Specifications

| Resin Type | Monomix dT(20) |
|-------------------------------------|--|
| Base & Support Matrix | Hydrophilic polymethacrylate, Monomix 1000 Å |
| Particle Size D50 (µm) | 30 |
| Average Pore Size (Å) | 1000 |
| Functional Group | Oligo dT20-mer |
| Ligand density | \geq 2.1 mg dT20 / mL of resin |
| Binding Capacity | >2.0 mg mRNA per mL of resin |
| mRNA Binding Capacity (mg/ml resin) | ≥2.0 mg |
| Max Linear Flow Rate (cm/hr) | 1000 |
| Operating Temperature (°C) | 4 - 65 |
| pH Stability | 2-12 |
| Operating Pressure | ≤100 bar (10 MPa) |
| Mobile Phase Compatibility | Compatible with common salt buffers, organic/aqueous solutions (acetonitrile, ethanol, etc.) and common additives for mRNA purification |
| Long-term Storage | Store in 20% ethanol aqueous solution, 2-25°C. Do not freeze resin or column. |
| CIP | 0.1-0.5M NaOH. Recommend to start with 0.1M NaOH to prolong resin life |





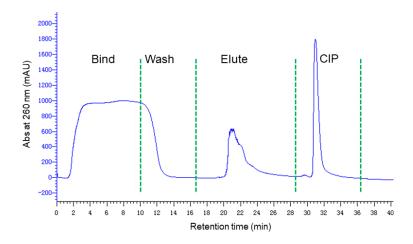


Sepax Monomix dT20 Affinity Resin COVID 19 mRNA vaccine candidate

Essential for mRNA commercial production, performance comparable to name brand

ANATOMY OF AN MRNA

5' cap



Coding region 3' UTR Poly(A) tail

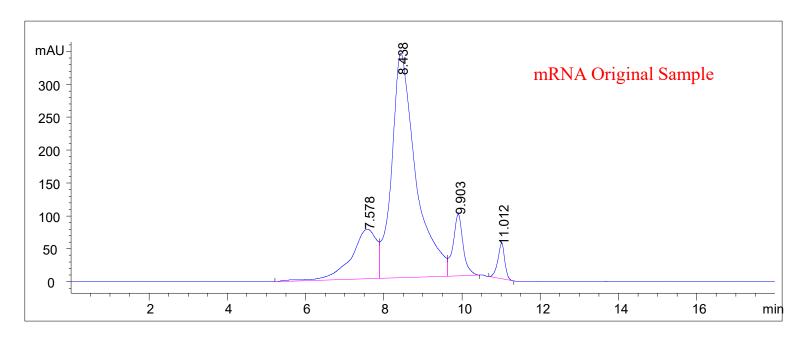
5' UTR

mRNA COVID-19 Vaccine Capture



mRNA Sample Analysis on SEC-1000

Column: SRT SEC-1000 5μm, 7.8 x 300 mm; Part Number: <u>215950-7830</u>; Mobile Phase: 150mM PB pH7.0; Flow Rate: 1 mL/min; Detector: UV 260 nm; Column Temperature: RT; Injection Volume: 5 μl; Sample: mRNA original sample (1mg/L); Pressure: 61bar; Instrument: HPLC

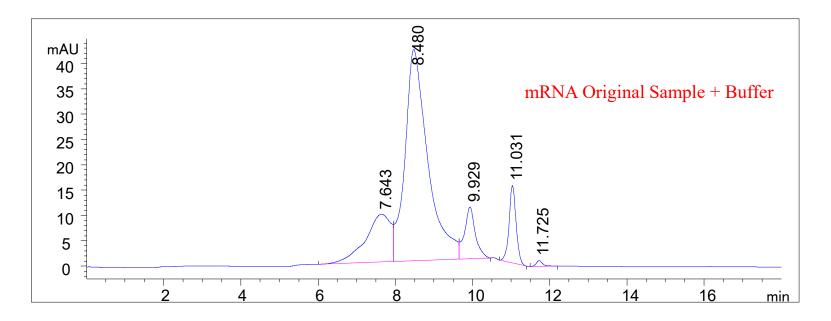


| # | RT | Area | Height | Width | Tailing | Area% |
|---|--------|---------|--------|--------|---------|--------|
| 1 | 7.578 | 3442.2 | 75.4 | 0.6659 | 1.965 | 16.826 |
| 2 | 8.438 | 14639.9 | 341.2 | 0.6416 | 0.579 | 71.562 |
| 3 | 9.903 | 1708 | 93.6 | 0.2658 | 1.034 | 8.349 |
| 4 | 11.012 | 667.6 | 54.2 | 0.1827 | 1.185 | 3.263 |



Sample + Buffer on SEC-1000

Column: SRT SEC-1000 5μm, 7.8 x 300 mm; Part Number: <u>215950-7830</u>; Mobile Phase: 150mM PB pH7.0; Flow Rate: 1 mL/min; Detector: UV 260 nm; Column Temperature: RT; Injection Volume: 5 μl; Sample: mRNA Original + Buffer (380ul dilute to 1ml); Pressure: 61bar

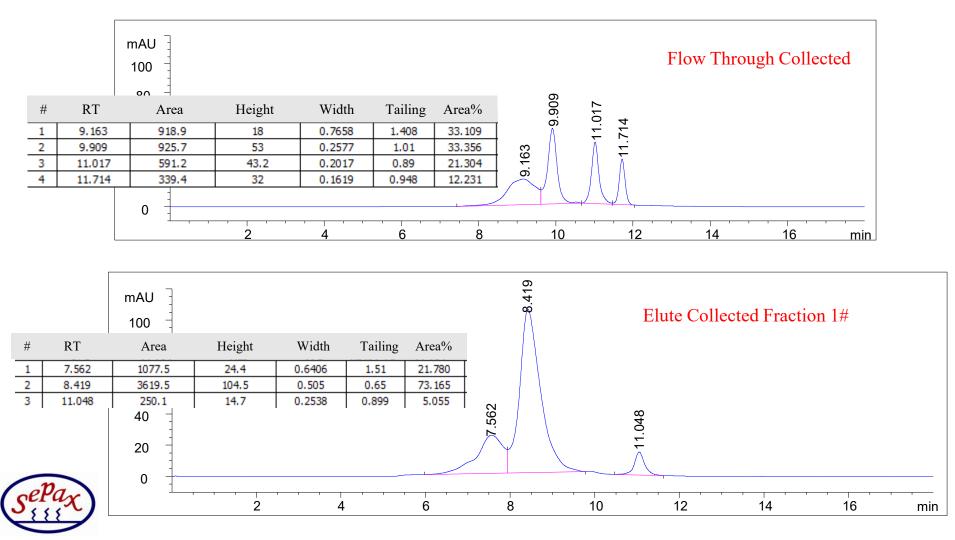


| # | RT | Area | Height | Width | Tailing | Area% |
|---|--------|-------|--------|--------|---------|--------|
| 1 | 7.643 | 427.5 | 9.4 | 0.6492 | 1.902 | 16.803 |
| 2 | 8.48 | 1723 | 41.7 | 0.5811 | 0.611 | 67.726 |
| 3 | 9.929 | 200.2 | 10.2 | 0.281 | 0.983 | 7.870 |
| 4 | 11.031 | 193.4 | 15.3 | 0.1899 | 0.902 | 7.601 |



Collected Fractions Analysis on SEC-1000

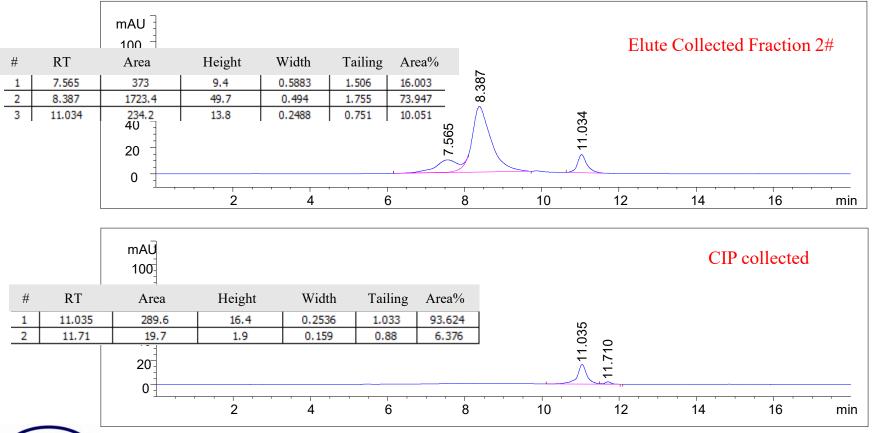
Column: SRT SEC-1000 5µm, 7.8 x 300 mm; Part Number: <u>215950-7830</u>; Mobile Phase: 150mM PB pH7.0; Flow Rate: 1 mL/min; Detector: UV 260 nm; Column temperature: RT; Injection Volume: 20µl; Sample: Collected Fractions; Pressure: 61bar



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Collected Fractions Analysis on SEC-1000

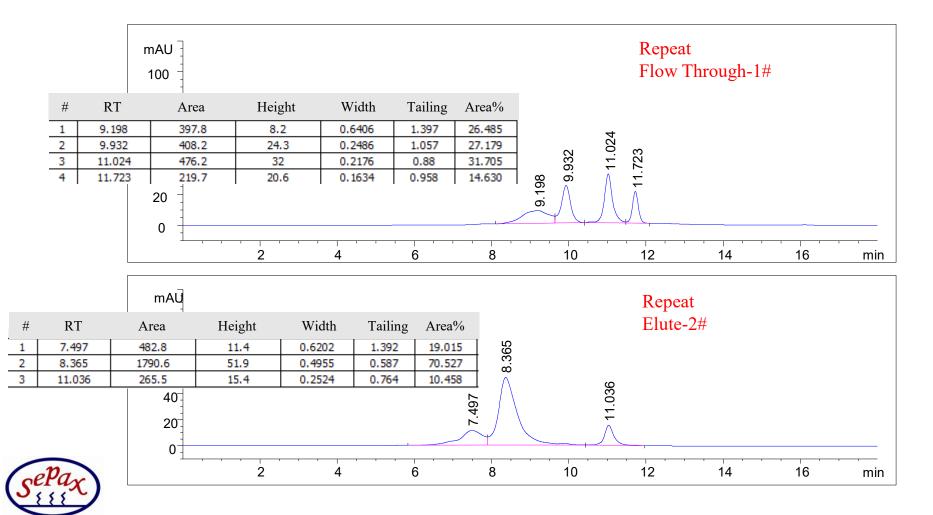
Column: SRT SEC-1000 5µm, 7.8 x 300 mm; Part Number: <u>215950-7830</u>; Mobile Phase: 150mM PB pH7.0; Flow Rate: 1 mL/min; Detector: UV 260 nm; Column Temperature: RT; Injection Volume: 20 µl; Sample: Collected Fractions; Pressure: 61bar



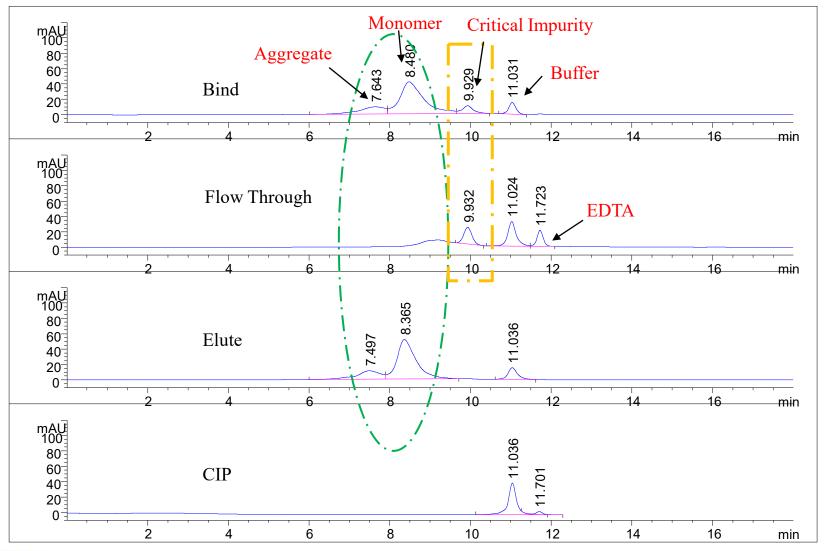


Collected Fractions Analysis on SEC-1000

Column: SRT SEC-1000 5 µm, 7.8 x 300 mm; Part Number: <u>215950-7830</u>; Mobile Phase: 150mM PB pH7.0; Flow Rate: 1 mL/min; Detector: UV 260 nm; Column Temperature: RT Injection Volume: 20µl; Sample: Collected Fractions; Pressure: 61bar



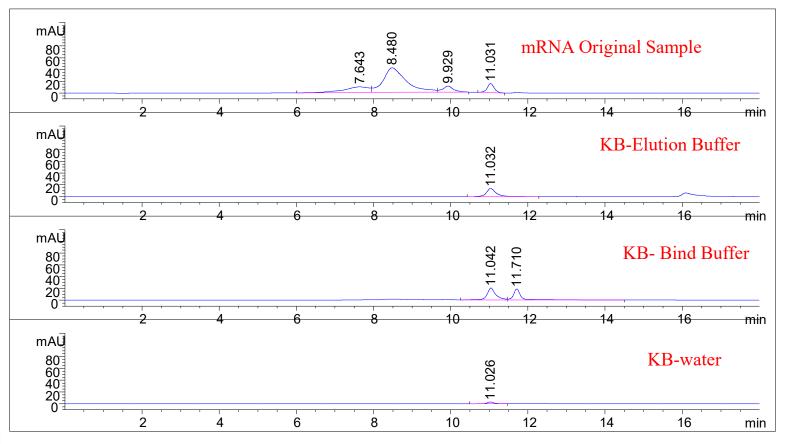
Fractions Overlays





Buffer Overlays

Column: SRT SEC-1000 5µm, 7.8 x 300 mm; Part Number: <u>215950-7830</u>; Mobile Phase: 150mM PB pH7.0; Flow Rate: 1 mL/min; Detector: UV 260 nm; Column Temperature: RT Injection Volume: 20µl; Sample: Blank; Pressure: 61 bar







mRNA Separation on SEC and RP

SRT SEC-1000, 5 μm, 1000 Å, 7.8 x 300 mm Part Number: 215950-7830

Proteomix RP-1000, 5 μm, 1000 Å, 2.1 x 100 mm Part Number: 465950-2110

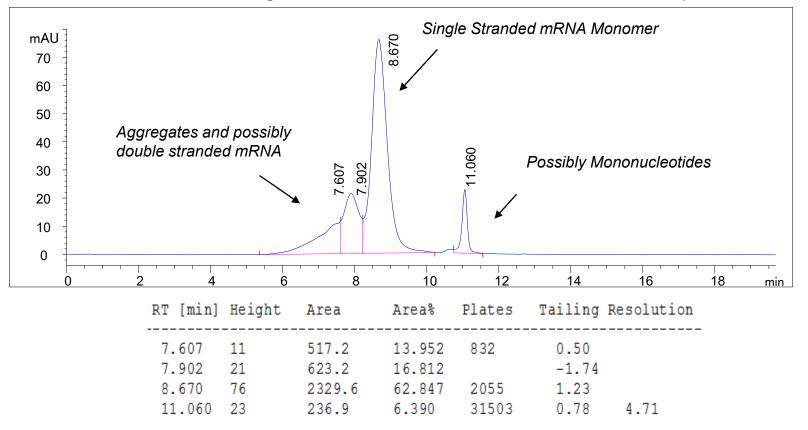
Sample: mRNA (Single-stranded, 1000 nucleotides, 300K ~ 600K Da) Concentration: 0.5 mg/mL Sample solution: Water (DEPC) Sample pH: Neutral Storage: -20 °C Impurity: Aggregates, fragments, double strand impurities

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mRNA-1 on SRT SEC-1000

Column: SRT SEC-1000, 5 μ m, 1000 Å, 7.8 x 300 mm (<u>215950-7830</u>); Mobile Phase: 150 mM PB, pH 7.0; Flow Rate: 1.0 mL/min; Detector: UV 260 nm; Column Temperature: 25 °C; Injection Volume: 5 μ L, Sample: mRNA-1 0.5 mg/mL

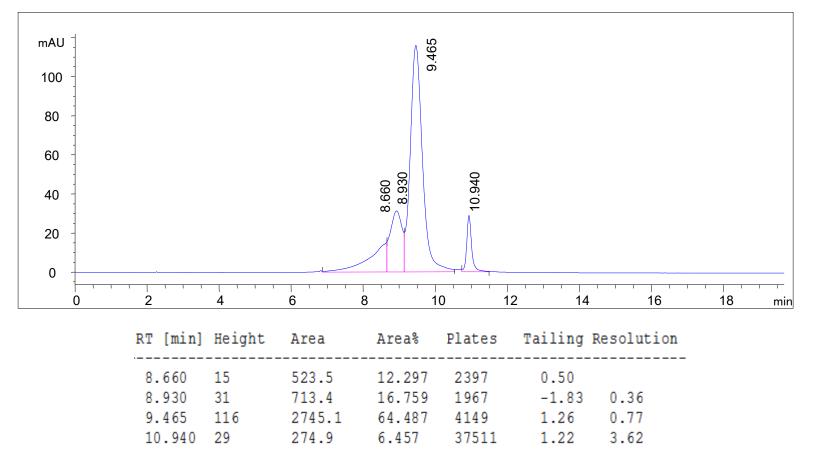


RP can be used as orthogonal method for further characterization of each SEC peak.



mRNA-1 on SRT SEC-2000

Column: SRT SEC-2000, 5 μ m, 1000 Å, 7.8 x 300 mm (<u>215980-7830</u>); Mobile Phase: 150 mM PB, pH 7.0; Flow Rate: 1.0 mL/min; Detector: UV 260 nm; Column Temperature: 25 °C; Injection Volume: 5 μ L, Sample: mRNA-1 0.5 mg/mL

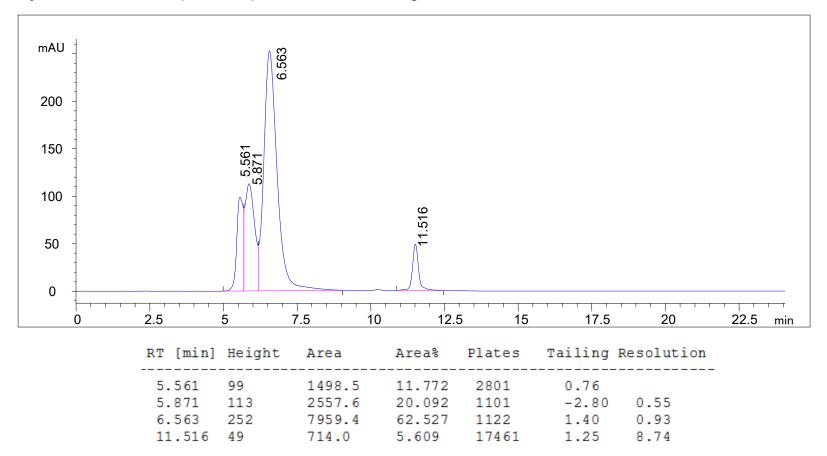


PI)()



mRNA-1 on SRT-C SEC-500

Column: SRT-C SEC-500, 5 μm, 500 Å, 7.8 x 300 mm (<u>235500-7830</u>); Mobile Phase: 150 mM PB, pH 7.0; Flow Rate: 1.0 mL/min; Detector: UV 260 nm; Column Temperature: 25 °C; Injection Volume: 5 μL, Sample: mRNA-1 0.5 mg/mL

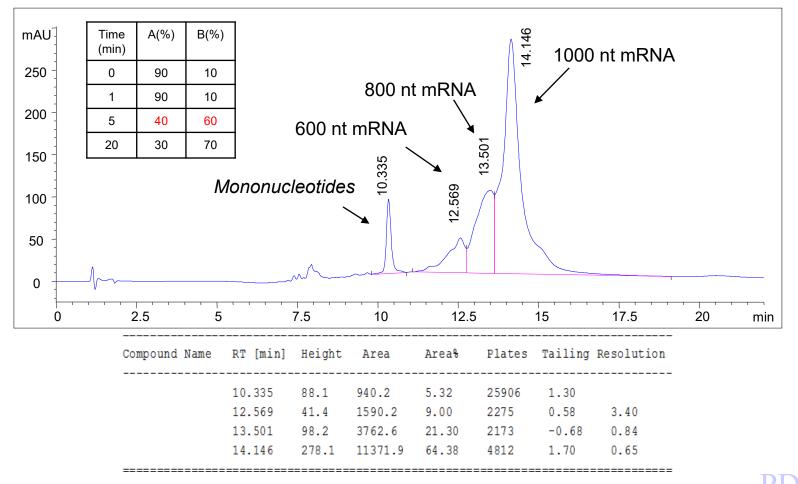


 $P(\mathbf{y})$



mRNA-2 Analysis on Proteomix RP-1000

Column: Proteomix RP-1000, 5 μm, 1000 Å, 2.1 x 100 mm (<u>465950-2110</u>); Mobile Phase: *A:* 100 mm TEAA *B:* 100 mm TEAA / 25% ACN; Flow Rate: 0.3 mL/min; Detector: UV 260 nm; Column Temperature: 50 °C; Injection Volume: 10 μl; Sample: mRNA-2; Pressure: 95 bar





SEPAX LITERATURE REFERENCE

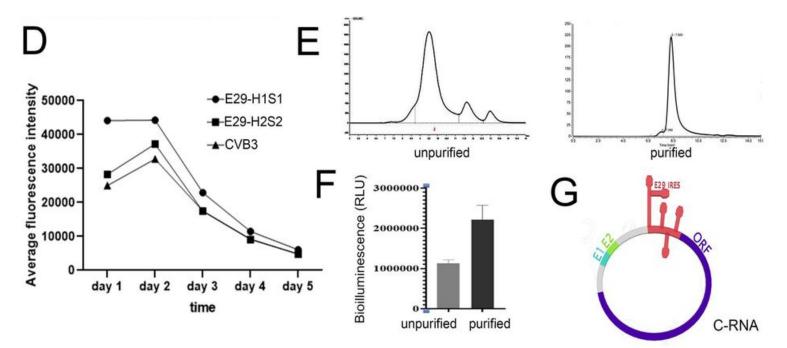
Circular mRNA Purification on SRT SEC 1000

Intratumoral Delivered Novel Circular mRNA Encoding Cytokines for Immune Modulation and Cancer Therapy

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Literature Reference: LR2022021104 Circular mRNA purification on SRT SEC 1000

Intratumoral Delivered Novel Circular mRNA Encoding Cytokines for Immune Modulation and Cancer Therapy



(D) Quantification of Fig.1C by FACS; (E) Diagram of HPLC shows the elution of circular mRNA before (left) and after (right) HPLC-SEC purifications; (F) Luciferase activities were measured 24 h post transfection of C-RNA with or without HPLC-SEC purifications; (G) Scheme of C-RNA.

Sepax PREP LC Column, SRT SEC-1000, 5um, 1000 A, 30 x 300 mm Part Number: 215950-30030

Curemed Biopharma Technology

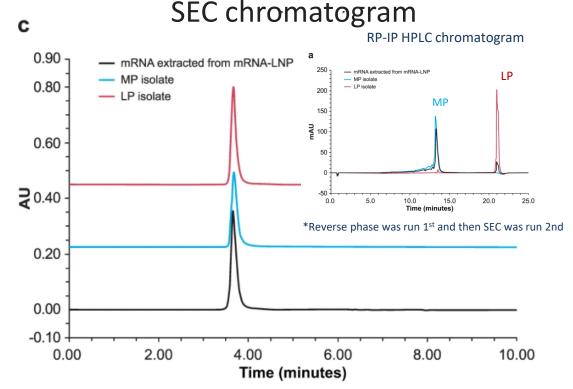
Yang, Jiali, et al. "Intratumoral Delivered Novel Circular mRNA Encoding Cytokines for Immune Modulation and Cancer Therapy." bioRxiv (2021). <u>https://doi.org/10.1101/2021.11.01.466725</u>

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Literature Reference: mRNA-LNP (Lipid Nanoparticle) on Sepax Analytical SEC

A novel mechanism for the loss of mRNA activity in lipid nanoparticle delivery systems

| Column | Zenix SEC-300 4.6x 150mm Part Number: 213300-4615 |
|------------------------|---|
| Mobile Phase | 100mM Tris acetate/2.5mM EDTA pH 8 |
| Flow Rate/Detection | 0.25 mL/min, UV 260nm |
| Instrument | Waters H-Class UPLC |
| Sample Notes | mRNA extracted from formulated mRNA-LNP |
| Length of Sample | ~2500-3000 nucleotides |
| mRNA: Lipid prep | mRNA was extracted from the mRNA-LNP formulation by IPA precipitation. (IPA and then NH4-Acetate) Dry-vacuo and then resuspended in RNase-free H2O |
| MP and LP prep | RP-IP HPLC on extracted mRNA from LNP's and fractionated. Generating purified MP and LP fractions MP and LP fractions re-injected onto RP- IP HPLC and SEC |



SEC was used to rule out if tertiary mRNA structures (i.e. aggregates) were the cause of the Late Peak

The SEC profile of the extracted mRNA vs. MP (main peak) vs. LP (late peak) were identical, thus eliminating aggregation as the origin of the late peak. This implicates other chemical reactions occurring to cause the generation of the late peak.



ModeRNA Therapeutics

Packer, Meredith, et al. "A novel mechanism for the loss of mRNA activity in lipid nanoparticle delivery systems." Nature communications 12.1 (2021): 1-11.

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Order Information

SRT SEC-1000, 5 μm, 1000 Å, 7.8 x 300 mm Part Number: 215950-7830

Proteomix RP-1000, 5 μm, 1000 Å, 2.1 x 100 mm Part Number: 465950-2110

Monomix dT20 Affinity Resin 1 mL, 5 mL resin Part Number: 283030950-0000

Monomix dT20 Affinity Resin 4.2 mL cartridge Part Number: 283030950-750100

Monomix dT20 Affinity 2.1 x 50 mm PEEK Part Number: 283030950P-2105

Monomix dT20 Affinity 4.6 x 50 mm PEEK

Part Number: 283030950P-4605

Resins and Columns are available for all your purification needs as well.

For Quotes or orders: <u>sales@sepax-tech.com</u> Phone: 1-877-SEPAX-US



For Technical Questions/ Method Development/ Services/ Seminar Requests: techsupport@sepax-tech.com Follow Us Website: www.sepax-tech.com LinkedIn: Sepax Technologies Facebook: @Sepaxtech

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